

## Physicochemical analysis of casein solubility in water-ethanol solutions

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**Abstract** – Due to the change in the apparent acid dissociation constant, when ethanol was added to water solutions of an industrially produced spray-dried sodium caseinate, the solution's pH increased proportionally to the reciprocal relative dielectric constant ( $1/\epsilon$ ). The pH increased by 1 unit when the ethanol volume fraction reached about 0.60. The casein solubility profile at 20 °C as a function of pH (3–8) and the ethanol volume fraction (0–0.75) was quite well represented by the equation describing the ionisation process. The pH inflection point and the slope coefficient were approximately linearly related to  $1/\epsilon$ . The evolution of the electrostatic surface potential ( $\psi_0$ ) and the zeta potential ( $\zeta$ ) explained the changes in casein solubility as a function of pH and the dielectric constant of the solvent. The increase in  $\psi_0$  by  $37 \pm 5.2$  mV or in  $\zeta$  by  $8.6 \pm 1.4$  mV raised casein solubility from 10 to 90%.

**casein / solubility / ethanol / dielectric constant / electrostatic potential**

**摘要** – 水-乙醇溶液中酪蛋白溶解性的理化分析。将乙醇加入到由喷雾干燥生产的酪蛋白酸钠水溶液后，由于表观酸解常数的变化，溶液的 pH 增加与介电常数的倒数 ( $1/\epsilon$ ) 成正比。当加入乙醇的体积分数达到 0.6 时，溶液的 pH 增加了 1 个单位。在 20 °C 时，酪蛋白的溶解性与 pH (3–8) 和乙醇体积分数 (0–0.75) 之间的函数关系可以很好地用电离方程描述。pH 拐点和斜率系数与  $1/\epsilon$  基本上呈线性关系。表面静电势 ( $\psi_0$ ) 和 zeta 电势 ( $\zeta$ ) 的变化进程可以很好地解释酪蛋白的溶解性与 pH 和溶剂的介电常数之间函数变化。当  $\psi_0$  增加到  $37 \pm 5.2$  mV 或者  $\zeta$  增加到  $8.6 \pm 1.4$  mV，酪蛋白的溶解性由 10% 增加达到 90%。

**酪蛋白 / 溶解性 / 乙醇 / 介电常数 / 静电势**

**Résumé** – Analyse physicochimique de la solubilité de la caséine dans des solutions d'éthanol-eau. En raison du changement de la constante apparente de la dissociation acide, l'ajout d'éthanol aux solutions aqueuses d'un caséinate de sodium industriel séché par pulvérisation, provoque une augmentation du pH des solutions inversement proportionnelle à la constante diélectrique ( $1/\epsilon$ ). Le pH augmente d'une unité quand la fraction volumique d'éthanol atteint environ 0.6. Le profil de la solubilité de la caséine à 20 °C, pour des pH de 3 à 8 et pour la fraction volumique d'éthanol entre 0 et 0.75, est assez bien représenté par une équation qui décrit le processus d'ionisation. Le point d'inflexion et la pente sont approximativement linéairement liés à  $1/\epsilon$ . L'évolution du potentiel électrostatique de surface ( $\psi_0$ ) et du potentiel zeta ( $\zeta$ ) explique les changements de la solubilité de la caséine en fonction du pH et de la constante diélectrique du solvant. L'augmentation du  $\psi_0$  de  $37 \pm 5.2$  mV ou du  $\zeta$  de  $8.6 \pm 1.4$  mV élève la solubilité de la caséine de 10 à 90 %.

**caséine / solubilité / éthanol / constante diélectrique / potentiel électrostatique**

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## 1. INTRODUCTION

Casein, being a mixture of  $\alpha_{S1}$ -,  $\alpha_{S2}$ -,  $\beta$ - and  $\kappa$ -casein fractions, represents an important and valuable source of ingredients due to its specific nutritional and functional properties [42, 43, 63, 73].

The solubility of main casein fractions in water is a function of the pH, temperature, dielectric constant, ionic strength and ionic environment (calcium and phosphate content) [7, 11, 16, 19, 55, 56, 58, 59].

Since the introduction in the XIXth century of the alcohol test for grading milk, several articles have been published on milk stability in ethanol solutions as a function of both alcohol concentration and Ca, Mg, Na, K, phosphate and citrate ion content, as well as pH, rennet, microbial contamination and genetic variants of casein fractions [8, 14, 28, 31–36, 40, 45, 54, 55, 57, 65, 68, 70, 76, 77].

Alcohol can also be used in the preparation and the purification of individual milk protein fractions [30, 39, 78, 80].

The stability of sodium caseinate in alcohol solutions is an important property for cream liqueur preparations [3–6, 15, 41, 46–48, 50].

Important differences in the secondary structure of milk proteins have been observed in alcohol – water solutions [9, 17, 18].

The aim of this work was to analyse from a physicochemical point of view the combined effects of alcohol concentration and pH on the solubility, the electrostatic surface potential and the electrostatic surface energy of sodium caseinate in water-ethanol mixtures.

## 2. MATERIALS AND METHODS

Industrially produced, spray-dried sodium caseinate (Armor Proteines, 35466 Saint Brice en Cogles, France) was reconstituted in deionised water at 50 °C using a laboratory mixer, cooled to 20 °C in a water bath and left overnight at room temperature to achieve a complete rehydration. To prevent bacterial growth and limit plasmin activity, 0.1 g·L<sup>-1</sup> NaN<sub>3</sub> and 0.1 g·L<sup>-1</sup> soybean

trypsin inhibitor were added [11]. The pH of the solutions was adjusted to values in the 2.5–8.0 range with 0.5 mol·L<sup>-1</sup> HCl or 0.5 mol·L<sup>-1</sup> NaOH and allowed to equilibrate for 1 h at room temperature. Ethanol was slowly added at 20 °C with continuous mixing, to reach the final ethanol volume fraction of up to 0.75. The solutions were then kept for 1 h at 20 °C and continuously stirred in a water bath. The casein concentration in final solutions was 10 g·L<sup>-1</sup>. Soluble caseins were separated from aggregates by centrifugation at 2000× *g* for 30 min. Casein content in solutions and supernatants was estimated from nitrogen determinations.

The total solids content of the caseinates was estimated after drying to a constant weight at 102 °C. Total nitrogen (TN), non-protein N (NPN) and non-casein N (NCN) were measured using Kjeldahl techniques with 6.38 as a nitrogen to protein conversion coefficient [37, 38]. Calcium (Ca), sodium (Na), magnesium (Mg) and potassium (K) concentrations were measured with a Varian SpectrAA 55B atomic absorption spectrometer. The pH of water and ethanol solutions was measured with a Radiometer pHM meter equipped with a combination electrode. Calibration was performed with buffers in aqueous solution. The relative permittivities of aqueous ethanol solutions were calculated according to Åkerlöf [1], Harvey and Prausnitz [27] and Smith et al. [64].

The ionic strength was calculated according to the relation:

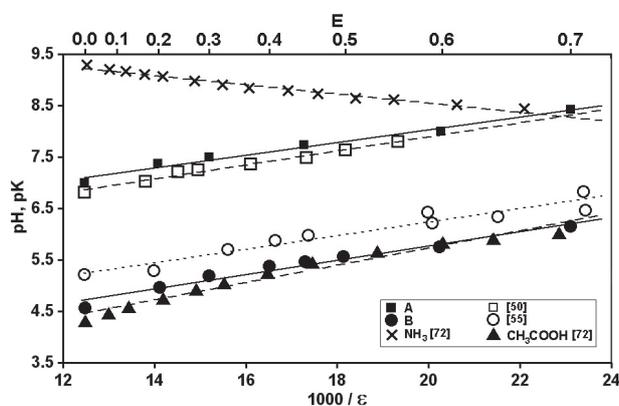
$$I = 0.5 \sum m_i z_i^2 \quad (1)$$

where: (*m*) denotes the molarity and (*z*) the valence of the dissolved ions.

The activity coefficient ( $\gamma$ ) of the ions was calculated by the equation [74]:

$$\gamma = \exp \left[ -42 \times 10^5 |z_+ z_-| \sqrt{\frac{I_T}{(\epsilon T)^3}} \right] \quad (2)$$

where: (*z*<sub>+</sub>) and (*z*<sub>-</sub>) are the valences of the positive and negative ions present in the solution, ( $\epsilon$ ) is the relative dielectric constant of the solvent and (*T*) is the absolute temperature.



**Figure 1.** The pH shift of sodium caseinate in water / ethanol solutions as a function of the reciprocal relative dielectric constant ( $\epsilon$ ): E = ethanol volume fraction. (■) – 10 g·L<sup>-1</sup> sample (A) – initial pH 7.0; (□) – 30 g·L<sup>-1</sup> sodium caseinate [50]; (●) – 10 g·L<sup>-1</sup> sample (B) = sample A with initial pH adjusted to 4.6; (○) – minimum ethanol stability of sodium caseinate [55]; (▲) – apparent pK of acetic acid [72]; (×) – apparent pK of ammonium [72]. Solid, broken and dotted lines represent equation (1) with coefficients A and 1/B given in Table I.

The hypothetical concentration of the ionised acidic ( $z_-$ ) or basic ( $z_+$ ) amino acids in mol per mol of protein, as a function of pH, was estimated by applying the relations [74]:

$$z_- = \frac{A_a}{1 + 10^{[pK_a - pH + \log(\gamma)]}} \quad (3)$$

$$z_+ = \frac{A_b}{1 + 10^{[pH - pK_a - \log(\gamma)]}} \quad (4)$$

where: ( $A_a$ ) and ( $A_b$ ) are, respectively, the concentrations, in mol per mol of protein, of a given acidic or basic amino acid and  $pK_a$  is the intrinsic acid dissociation constant, i.e. the pH level at which the dissociation is exactly 50% for the ionic strength  $I = 0$ ; ( $\gamma$ ) is the activity coefficient.

### 3. RESULTS AND DISCUSSION

#### 3.1. Composition

The sodium caseinate used in this study contained over 930 g·kg<sup>-1</sup> dry matter and over 960 g·kg<sup>-1</sup> casein in total protein. It is consistent with the results of Muir and

Dalglish [48] obtained for industrial sodium caseinates. The sodium content of about 14.6 g·kg<sup>-1</sup> was in agreement with the results of Towler [69–71]. The potassium and the magnesium contents were 2.5 g·kg<sup>-1</sup> and 0.3 g·kg<sup>-1</sup>, respectively. The calcium level (1.5 g·kg<sup>-1</sup>) was close to that found by Muir and Dalglish [48] for sodium caseinates from various suppliers.

#### 3.2. pH shift

For ethanol volume fractions up to 0.75, the pH of sodium caseinate solutions rose linearly with the increase in the reciprocal relative dielectric constant ( $\epsilon$ ) according to the relation (Fig. 1):

$$pH = A + B/\epsilon + \text{Err} \quad (5)$$

where: A is the intercept or the pH level for the hypothetical  $\epsilon = \infty$  and the slope coefficient B indicates the hypothetical pH increase for unitary increase in  $1/\epsilon$ ; Err is the standard error term following a normal distribution. More evocative is the reciprocal value of the coefficient B, which indicates the change in  $1/\epsilon$  causing the pH or the apparent pK to change by 1 unit:

$$A + B/\epsilon = A + B/\epsilon_w + 1, \text{ so } 1/\epsilon = 1/\epsilon_w + 1/B$$

**Table I.** Intercept (A), slope (B), standard error (Err) and correlation coefficient ( $R^2$ ) from Equation (5) expressing the relation between the pH or the apparent pK and the reciprocal dielectric constant ( $1/\epsilon$ ) presented in Figure 1. N = number of experimental points.

Sample	A	B	Err	$R^2$	N	pH <sub>1</sub> or pK <sub>1</sub>	E <sub>1</sub>
Sample (A)	5.75 ± 0.27	123 ± 8.1	0.079	0.934	6	8.19	0.63
Sample (B)	3.00 ± 0.17	139 ± 9.8	0.088	0.971	8	5.73	0.57
Pierre [55]	3.56 ± 0.23	132 ± 12.3	0.141	0.934	10	6.23	0.58
CH <sub>3</sub> COOH [72]	2.35 ± 0.17	209 ± 6.9	0.113	0.967	12	5.46	0.51
O'Kennedy et al. [50]	5.17 ± 0.11	137 ± 6.9	0.044	0.984	8	7.86	0.58
NH <sub>3</sub> [72]	10.3 ± 0.06	-88.9 ± 3.7	0.039	0.976	15	11.46	0.71

pH<sub>1</sub> or pK<sub>1</sub> shows the pH or pK level for the reciprocal relative dielectric constant  $1/\epsilon = (1/\epsilon_w + 1/B)$ , where  $\epsilon_w = 80.2$  at 20 °C is the relative dielectric constant of water.

E<sub>1</sub> = ethanol volume fraction corresponding to pH<sub>1</sub> or pK<sub>1</sub>.

where  $\epsilon_w = 80.2$  at 20 °C is the relative dielectric constant of water.

The values of A and B are given in Table I.

The sodium caseinate samples examined in this study gave very similar tendencies in  $1/B$  to those calculated from the results of Pierre [55] and O'Kennedy et al. [50]. The apparent pK change in the acetic acid [72] was also close to the values obtained for different samples of sodium caseinate. Contrary to carboxyl groups, the apparent pK of amino groups decreases with the increased in the reciprocal relative dielectric constant [72]. The pK reduction rate of the amino groups was about 20% lower than that of the carboxyl groups.

When the reciprocal dielectric constant ( $1/\epsilon$ ) increased from its initial level ( $1/80.2 = 1.25 \times 10^{-2}$ ) by  $1/B$  from equation (1), the pH or the apparent pK of carboxyl groups increased by one unit to pH<sub>1</sub> or pK<sub>1</sub> (Tab. I). This happened when the ethanol volume fraction reached 0.5 to 0.7 for different samples and groups (E<sub>1</sub> in Tab. I).

The phenomenon of pH or pK changes is due to the evolution of the electrostatic energy when an ion is transferred from one solvent to another one, both characterised by the relative dielectric constants ( $\epsilon_1$ ) and ( $\epsilon_2$ ). According to Max Born's model of the electrostatic work (W in joules J) required

to transfer an ion of a radius (r) and the charge (z) is equal to [72]:

$$W = e^2 z^2 (1/\epsilon_1 - 1/\epsilon_2) / (8 \pi r \epsilon_0) \quad (6)$$

where: e = elementary electronic charge =  $1.602 \times 10^{-19}$  C,  $\pi = 3.14$ ,  $\epsilon_0 = 8.854 \times 10^{-12}$  J<sup>-1</sup>·C<sup>2</sup>·m<sup>-1</sup> = dielectric permittivity of vacuum.

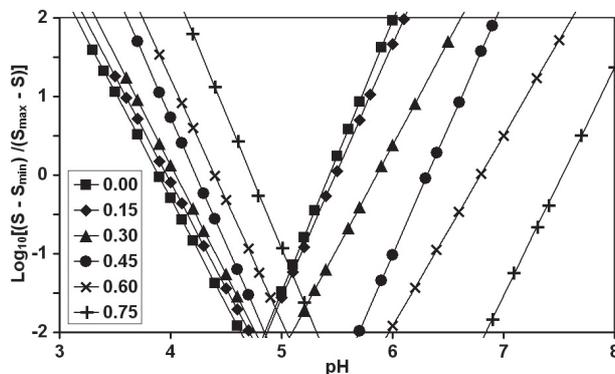
For casein monomers at pH 7 with a radius of around 4.5 nm [66], the electrostatic work (W) is about 30 kJ·mol<sup>-1</sup> for the passage from water to 60% ethanol, which is the average alcohol concentration for pH<sub>1</sub>, calculated for samples A and B and those of Pierre [55] and O'Kennedy et al. [50].

### 3.3. Protein solubility

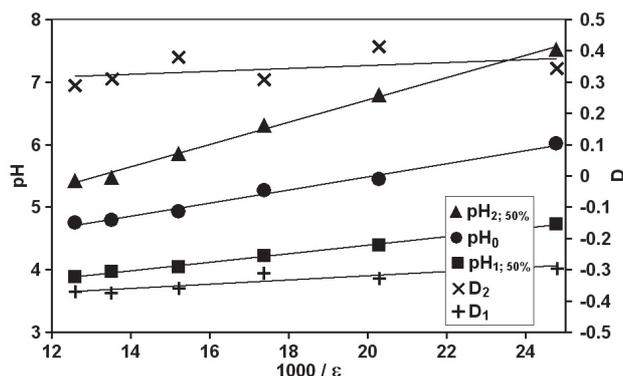
The solubility profile (Fig. 2) below and above the isoelectric point follows the Linderstrøm-Lang equation (7), representing the ionisation process [32–34]:

$$\text{Log}_{10}[(S - S_{\min})/(S_{\max} - S)] = (\text{pH}_{50\%} - \text{pH})/D \quad (7)$$

where: pH<sub>50%</sub> is the pH inflexion point or the pH corresponding to 50% casein solubility; D is the slope of the straight line in pH versus  $\text{Log}_{10}[(S - S_{\min})/(S_{\max} - S)]$  coordinates,  $S_{\min}$  and  $S_{\max}$  are, respectively, the minimal and maximal casein solubility



**Figure 2.** The effect of pH (3–8) and ethanol volume fraction (0–0.75) on the solubility profile of 10 g·L<sup>-1</sup> sodium caseinate solutions. Solid lines represent Equation (3) with coefficients pH<sub>1</sub> and D presented in Figure 3 and Table II.



**Figure 3.** Evolution of pH corresponding to 50% casein solubility (pH<sub>1,50%</sub> and pH<sub>2,50%</sub>) and coefficient D from Equation (3) as a function of the reciprocal relative dielectric constant (1/ε) for the pH range below (pH<sub>1</sub> and D<sub>1</sub>) and over (pH<sub>2</sub> and D<sub>2</sub>) the minimal solubility point (pH<sub>0</sub>). Solid lines represent Equation (1) with coefficients A and B given in Table II.

and S is the solubility at a given pH and ethanol concentration.

Equation (7) can also be presented in the form:

$$S = \frac{S_{\min} + S_{\max} \times 10^{(pH_{50\%} - pH)/D}}{1 + 10^{(pH_{50\%} - pH)/D}} \quad (8)$$

For the pH range below the minimal solubility point (pH<sub>0</sub>), pH<sub>1,50%</sub> from equations (7 or 8) increased linearly between 3.7 and 4.6

with the rise in 1/ε (Fig. 3, Tab. II). For pH values over the minimal solubility point, pH<sub>2,50%</sub> rose much more (5.2 to 7.4). The lowest level of casein solubility (pH<sub>0</sub>) in water occurred at pH 4.7 and increased linearly with the rise in 1/ε (Fig. 3). The low solubility zone broadened with the increase in ethanol concentration. A similar tendency for casein solubility in water and in up to 66% ethanol was observed by Zittle and Pepper [79] and by Pierre [55]. The variation in the coefficient D from equation (8) is

**Table II.** Intercept (A), slope (B), standard error (Err) and correlation coefficient ( $R^2$ ) from Equation (5) expressing the linear relations between the pH and coefficient (D) from Equation (7) and the reciprocal dielectric constant ( $1/\epsilon$ ) presented in Figure 3. Number of experimental points  $N = 6$ .

	A	B	Err	$R^2$
$\text{pH}_{1,50\%}$	$3.03 \pm 0.03$	$68 \pm 1.6$	0.017	0.998
$\text{pH}_{2,50\%}$	$3.16 \pm 0.10$	$178 \pm 5.6$	0.058	0.996
$\text{pH}_0$	$3.39 \pm 0.10$	$104 \pm 5.7$	0.060	0.988
$D_1$	$0.45 \pm 0.03$	$-6.7 \pm 1.5$	0.015	0.838
$D_2$	$-0.26 \pm 0.08$	$-4.8 \pm 4.6$	0.048	0.211

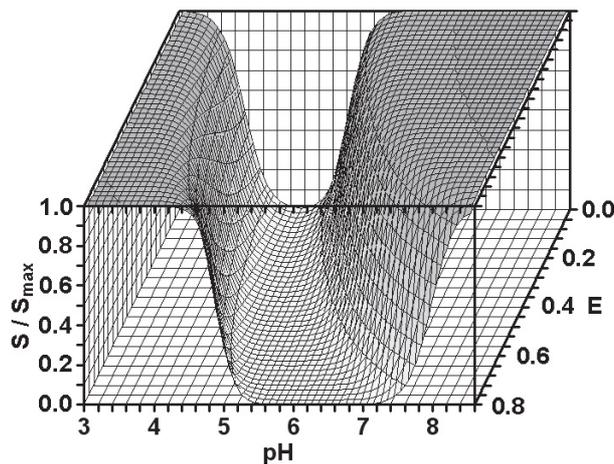
relatively small in comparison with the experimental error (Fig. 3). It is positive for the pH range below the minimal solubility point. For the pH range over the minimal solubility point, it is negative and on average does not change with the rise in the ethanol concentration. In absolute terms, there is no statistically significant difference between  $D_1$  and  $D_2$  and their average level is  $0.339 \pm 0.039$ .

Around  $\text{pH}_{50\%}$  points for pH values below and above the minimal solubility point, the casein solubility decreased or

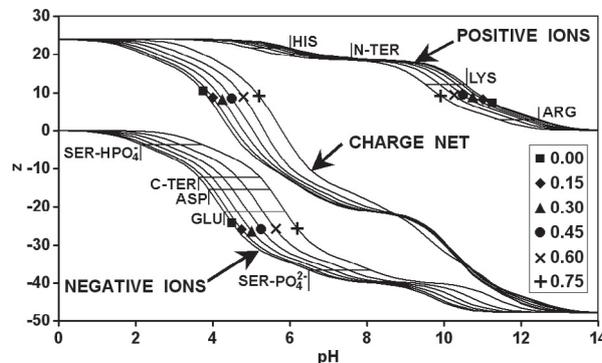
increased very rapidly with pH change. Thus, any slight imperfections in the estimation of pH or in the solubility cause quite a significant scattering of the coefficient D between different individual casein solubility curves as a function of pH for a given ethanol concentration (Fig. 3). Nevertheless, for the whole set of data, the regression line between the experimental ( $S_{\text{EXP}}$ ) and model ( $S_{\text{MOD}}$ ) values of casein solubility for  $N = 150$  experimental points gives a standard error of around 3%, which is quite satisfactory:

$$S_{\text{MOD}} = 0.1(\pm 0.35) + 0.996(\pm 0.005)S_{\text{EXP}}; \\ R^2 = 0.996; \text{Err} = 2.8; N = 150.$$

The casein solubility evolution (Fig. 4), calculated by equation (8) shows two solubility zones over 90%, limited by the isolines joining the points of pH 3.5 for water with pH 4.6 for a 0.8 ethanol volume fraction and pH 5.7 for water with pH 8.3 for a 0.8 ethanol volume fraction. The lower solubility zone (<10%) is limited by the isolines passing from pH 4.2 for water to pH 5.1 at 0.8 ethanol volume fraction and from pH 5.1 at 0% to pH 7.6 for 80% ethanol solutions. Between these relatively flat lower or higher solubility zones, there are two very steep zones of decreasing and



**Figure 4.** Relative casein solubility ( $S/S_{\text{max}}$ ) as a function of pH and ethanol volume fraction (E), calculated by Equation (8) with  $S_{\text{min}} = 0$ ,  $S_{\text{max}} = 100$  and with average  $\text{pH}_{50\%}$  and D coefficients calculated by the regression equations from Table II.



**Figure 5.** Evolution of the electrical charge ( $z$ ) of total casein monomers, as a function of pH and ethanol volume fraction (0 to 0.75), calculated by Equations (3) and (4) with  $\gamma = 1$  and pK taken from Walstra and Jenness [75]. Horizontal lines with amino acid name abbreviations representing their apparent pK evolution as a function of the ethanol volume fraction (0–0.75).

**Table III.** Intercept (A), slope (B), standard error (Err) and correlation coefficient ( $R^2$ ) from Equation (5) representing the isolines of 10% and 90% of casein solubility (Fig. 4) and expressing the linear relations between the pH and the reciprocal dielectric constant ( $1/\epsilon$ ). Number of experimental points  $N = 6$ .

	A	B	Err	$R^2$
$\text{pH}_{1;10\%}$	$3.46 \pm 0.004$	$62 \pm 0.2$	0.003	0.999
$\text{pH}_{2;10\%}$	$2.71 \pm 0.009$	$166 \pm 0.5$	0.007	0.999
$\text{pH}_{1;90\%}$	$2.59 \pm 0.006$	$74.6 \pm 0.3$	0.005	0.999
$\text{pH}_{2;90\%}$	$3.41 \pm 0.003$	$182 \pm 0.2$	0.002	0.999

increasing solubility with rising pH. In pH against ( $1/\epsilon$ ) coordinates (Fig. 4, Tab. III), the 10% and 90% isolines of casein solubility can be represented by equation (5).

### 3.4. Ionisation

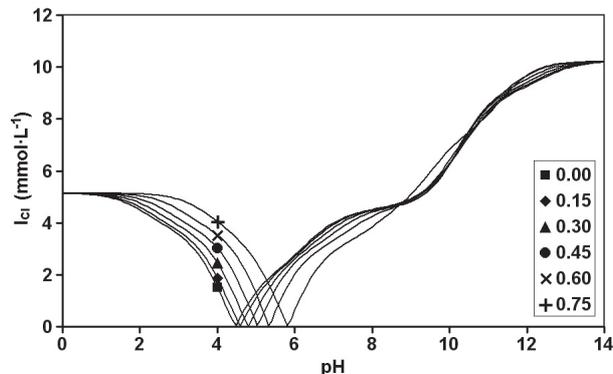
Protein solubility is in general a function of the intermolecular repulsion and attraction forces that depend mainly on the electrical charge, ionic strength and dielectric constant.

On average, the native casein contains  $190 \text{ mmol}\cdot\text{kg}^{-1}$  of histidine, 560 of lysine, 220 of arginine, 220 of aspartic acid, 740 of

glutamic acid and 260 of phosphate groups [75]. In total, casein contains  $1.35 \text{ mol}\cdot\text{kg}^{-1}$  of anionic groups against  $0.78 \text{ mol}\cdot\text{kg}^{-1}$  of cationic groups.

The content of  $\alpha_{S1-}$ ,  $\alpha_{S2-}$ ,  $\beta$ - and  $\kappa$ -caseins in total casein is on average equal, respectively, to 40.2, 10.5, 37.4 and 11.9% [66, 67]. On the basis of these proportions and on the known amino acid composition of the main casein fractions, we calculated the average content of the ionic groups in total casein.

The ionic casein groups, dissociated within the 4 to 9 pH range are  $\gamma$ - and  $\delta$ -carboxyl groups of the aspartic and glutamic acids and the amino group of histidine with pKs of 4.1, 4.6 and 6.4, respectively [55, 74, 75]. Typically for strong acids, the pK of the first  $\text{H}^+$  of the serine phosphate (pK 2.12) is only slightly dependent on the dielectric constant changes with ethanol concentration up 80% [72]. For the second  $\text{H}^+$  of phosphate esters, the pK 7.21 would increase as for other weak acids. The pK of amino groups decreases by 1 unit with the average increase in the ethanol volume fraction from 0 to 0.73 [72]. So, in 73% (v/v) ethanol the pK of the imidazole group of histidine would be 5.4, that of the  $\epsilon$ -amino group of lysine would decrease from 10.6 to 9.6 and of the guanidine group of arginine from 12.5 to



**Figure 6.** Evolution of the counter ions' ionic strength ( $I_{C1}$ ) in  $\text{mol}\cdot\text{L}^{-1}$ , as a function of pH and ethanol volume fraction (0 to 0.75), calculated by Equation (1), with  $m = 0.428 \text{ mmol}\cdot\text{L}^{-1}$  and the net charge ( $z$ ) taken from Figure 5.

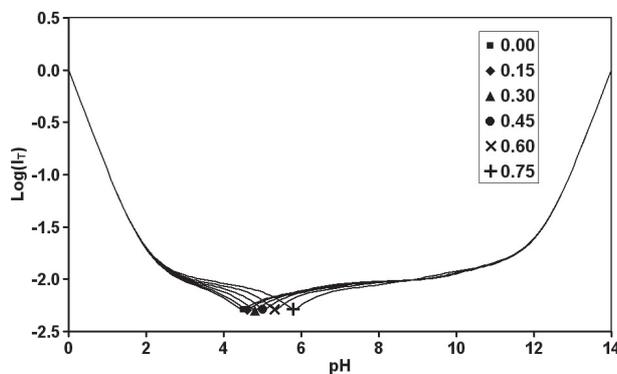
11.5. In water solution at pH 7 only histidine is at about 80% in its neutral form and at 20% in its protonated form. Lysine and arginine remain almost totally in their protonated forms up to pH 9. In 60% ethanol solutions the carboxyl groups would be 10 times less dissociated than in water. On the other hand, in 73% ethanol the amino groups would be 10 times less protonated, but among them only histidine can play a minor role in electrical charge modification of casein molecules.

For a given pH of casein water solution, when alcohol was added, the pH increased spontaneously, because of the apparent pK changed as an effect of the dielectric constant variation, but the electrical charge of a protein molecule or protein aggregate did not change (Fig. 5). On the other hand, for the same pH level for water and ethanol solutions, the charge differences can be very significant. For pH values between 0 and 3, the amino casein groups are >99.98% protonated. At pH 14 about 99.3% of amino groups are in their neutral form. At pH < 1, over 98.8% of the acidic casein groups are not dissociated. At pH > 11 in water solution and at pH > 12.5 in 75% ethanol solutions, over 99.5% of acidic groups are dissociated. Only at pH < 1 and over 13.5 is there practically only one sort of charge: positive at low pH and negative at high pH levels. At pH < 1, the average electrical charge ( $z$ ) of casein aggregates is +24 and

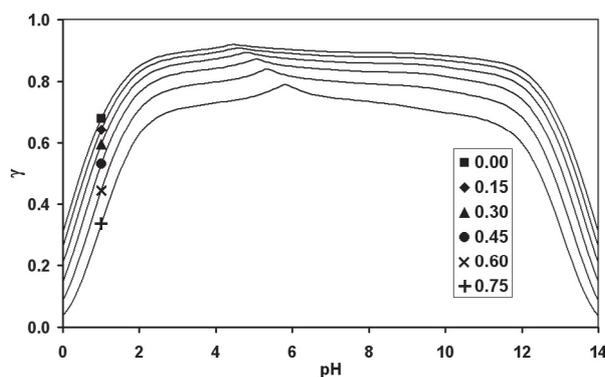
at pH 14 it is -48. Zero charge, i.e. equal numbers of positive and negative charges in a casein molecule or aggregate, is at pH 4.57 for water solutions and increases to pH 5.79 for 0.75 ethanol volume fractions. Swaisgood [66] found isoionic pH values for water solutions of different casein fractions of between 4.94 and 5.90. For native phosphocaseinate dissolved in water, 50% protein solubility was observed at pH 5.2 [21]. At pH 7.0 the average net charge of total casein would be -18.7 in water solution and -13.6 in 75% ethanol. Because of the opposite effect of alcohol on the pK of amino and acidic groups, for pH > 9 the net charge of casein molecules and aggregates would almost be independent of the ethanol concentration.

### 3.4.1. Ionic strength

The solution containing  $10 \text{ g}\cdot\text{L}^{-1}$  of sodium caseinate contains  $7.09 \text{ mmol}\cdot\text{L}^{-1}$   $\text{Na}^+$ ,  $0.71 \text{ K}^+$ ,  $0.42 \text{ Ca}^{2+}$  and  $0.14 \text{ Mg}^{2+}$ . The pH of this solution in water is 7.0. The minerals present in the solution represent an ionic strength of  $5 \text{ mmol}\cdot\text{L}^{-1}$ . The contribution of the casein to the ionic strength is as significant as that of the minerals already present. As the solution must be electrically neutral, the counter ions ( $\text{Cl}^-$  or  $\text{Na}^+$ ) have to be added to neutralise the protonated amino groups and the dissociated phosphoryl, carboxyl and thyrosyl groups. The ionic



**Figure 7.** Evolution of the logarithm of the total ionic strength ( $I_T$  in  $\text{mol}\cdot\text{L}^{-1}$ ), as a function of pH and ethanol volume fraction (0 to 0.75), calculated using Equation (1).



**Figure 8.** Evolution of the activity coefficient ( $\gamma$ ), as a function of pH and ethanol volume fraction (0 to 0.75), calculated by Equation (2) for  $T = 293$  K.

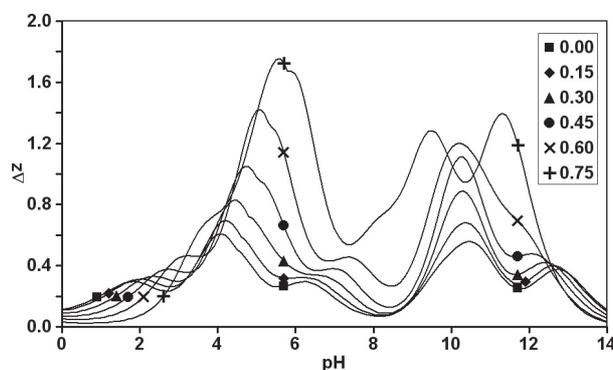
strength, which represents the counter ions, varied between 0 at the isoelectric point and  $5.1 \text{ mmol}\cdot\text{L}^{-1}$  at pH 0 and  $10.2 \text{ mmol}\cdot\text{L}^{-1}$  at pH 14 (Fig. 6). The greatest contribution to the total ionic strength ( $I_T$  in Fig. 7) was the ions that were added to the solution to reach very low or very high pH level.

### 3.4.2. Activity coefficient and electrical charge

The activity coefficient ( $\gamma$ ), calculated by Equation (2), varied between 0.04 and 0.92 (Fig. 8). It is at its highest at the isoelectric point and decreases sharply in very low and very high pH regions. The presence

of ethanol significantly reduces the activity coefficient.

When taking into account the activity coefficient ( $\gamma$ ) from Figure 8 in Equations (3) and (4), instead of  $\gamma = 1$ , the level of the ionised acidic and basic amino acids became different from that presented in Figure 5. This difference,  $\Delta z = z_{\gamma=1} - z_{\gamma}$  (Fig. 9), increased with the rise in the ethanol volume fraction. With the increasing ionic strength and decreasing dielectric constant, the activity coefficient decreased, so less energy is needed to remove a proton from a charged molecule, as the proton senses a smaller electrostatic potential [74]. The observed differences



**Figure 9.** Evolution of the difference ( $\Delta z = z_{\gamma=1} - z_{\gamma}$ ) between the net electrical charge ( $z$ ) of total casein monomers, calculated by Equations (3) and (4) for the activity coefficient  $\gamma = 1$  and for the values of  $\gamma$  taken from Figure 8, as a function of pH and ethanol volume fraction (0 to 0.75).

were greater around those pH levels close to the pK of the principal ionic amino acids, namely, glutamic and aspartic acids (pK = 4.3 and 3.9) and lysine and arginine (pK = 10.6 and 12.5). Because of the opposing effects of the dielectric constant and the ionic strength on the apparent pK of basic amino acids, quite wide differences in  $\Delta z$  were observed between 0.6 and 0.75 ethanol volume fractions. Close to the isoelectric point ( $4 < \text{pH} < 6$ ), the observed differences in  $\Delta z$  were larger than at the potential casein molecule charge within this pH range. However, as values grew further away from the isoelectric point these differences became relatively small, when the overall variation in  $z$  between +24 at pH 0 and -48 at pH 14 was taken into account.

### 3.5. Electrostatic surface and zeta potentials

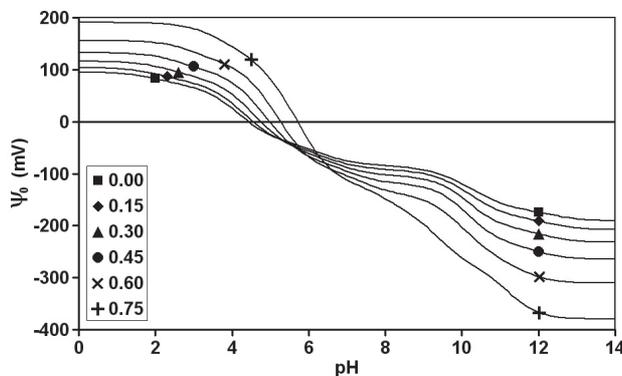
Casein molecules can be considered as spheres of an average radius  $r = 4.5$  nm [66]. Their average electrostatic surface potential ( $\psi_0$ ) can be estimated by the relation:

$$\psi_0 = ze / (4 \pi \epsilon_0 \epsilon r). \quad (9)$$

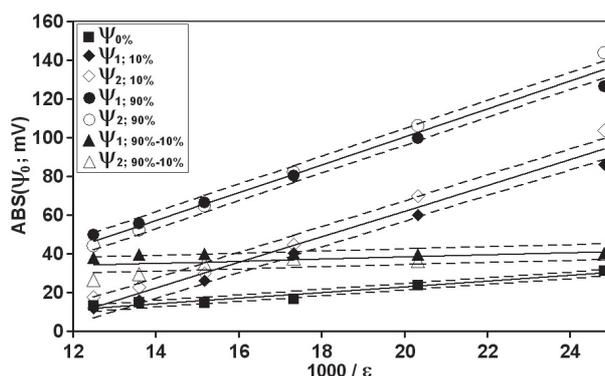
The  $\psi_0 = 0$  at the isoelectric point ( $\text{pH}_0$ ), which increases from about 4.5 for water solutions to 5.8 for 0.75 ethanol volume fractions (Fig. 10). For the  $\text{pH} < \text{pH}_0$  range,

the  $\psi_0$  is positive and for  $\text{pH} > \text{pH}_0$  it is negative. For casein dissolved in water or in 75% ethanol, the electrostatic surface potential at pH 0 is +95 mV and +191 mV, respectively. At pH 14 the  $\psi_0$  is, respectively, -189 and -376 mV.

The surface potential ( $\psi_0$ ) did not coincide strictly with the isoline  $\text{pH}_0$  from Figure 3. The surface potential, corresponding to the isoline  $\text{pH}_0$ , increased from 12 mV for water solutions to 30 mV for a 0.75 ethanol volume fraction (Fig. 11, Tab. IV). The absolute values of the electrostatic surface potentials  $\psi_{1;10\%}$ ,  $\psi_{2;10\%}$ ,  $\psi_{1;90\%}$ , and  $\psi_{2;90\%}$ , corresponding to 10% and 90% casein solubilities equal for the pH range below ( $_1$ ) and over ( $_2$ ) the minimal pH solubilities, were inversely proportional to the relative dielectric constant (Fig. 11). Within the standard error limits of ( $\pm 5$  mV), the electrostatic surface potential levels were similar for the corresponding isolines on both sides of the minimal casein solubility or minimal surface potential. The surface potential difference between 90% and 10% casein solubility was on average  $37 \pm 5.2$  mV, independently of the dielectric constant of the solvent. This means that on both sides of the isoelectric point, an increase in the surface potential of about 40 mV augments the casein solubility from 10% to 90%.



**Figure 10.** Evolution of the electrostatic surface potential ( $\psi_0$ ) of sodium caseinate monomers, as a function of pH and ethanol volume fraction (E), calculated by Equation (9) with the charge (z) calculated by Equations (3) and (4) and the activity coefficient ( $\gamma$ ) by Equation (2).



**Figure 11.** Absolute value of the electrostatic surface potential ( $\psi_0$ ) of casein monomers as a function of the reciprocal relative dielectric constant ( $1/\epsilon$ ) for the line  $pH_0$ , from Figure 3 ( $\psi_{0\%}$ ) and for the isolines  $pH_{1;10\%}$ ,  $pH_{2;10\%}$ ,  $pH_{1;90\%}$  and  $pH_{2;90\%}$  from Figure 4, corresponding to casein solubilities of 10% and 90% for the pH range below (1) and over (2) the minimal surface potential ( $\psi_0$ ). Solid lines represent Equation (5) with coefficients A and B given in Table IV and broken lines show standard deviation limits.

The electrostatic surface potential of protein molecules in solutions is partly screened by the counter ions present in the double layer. The potential at the double layer, being approximately equal to zeta potential ( $\zeta$ ), can be estimated by the relation [29]:

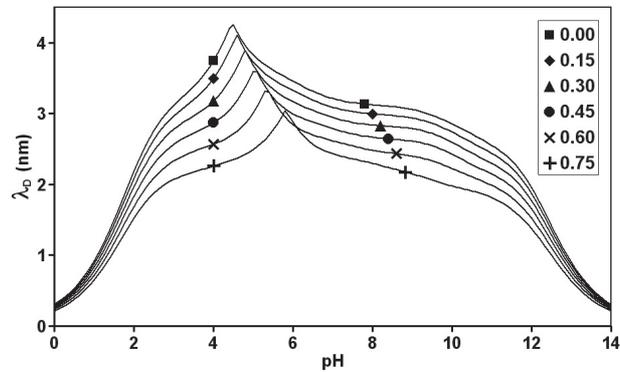
$$\zeta = \psi_0 \frac{r}{r + \lambda_D} \exp(-1) \quad (10)$$

where:  $\lambda_D$  is the Debye length or the nominal thickness of the electric double layer,

being a function of the relative dielectric constant of the solvent ( $\epsilon$ ), the absolute temperature (T) and the total ionic strength ( $I_T$ ), according to the equation [74]:

$$\lambda_D = \sqrt{\frac{\epsilon_0 \epsilon k_B T}{2 I_T N_A e^2}} \quad (11)$$

where:  $k_B = 1.381 \times 10^{-23} \text{ J}\cdot\text{K}^{-1}$  is the Boltzmann constant and  $N_A = 6.022 \times 10^{23}$  is the Avogadro number.



**Figure 12.** Evolution of the Debye length ( $\lambda_D$ ), as a function of pH and of ethanol volume fraction (0 to 0.75), calculated by Equation (11) for  $T = 293$  K.

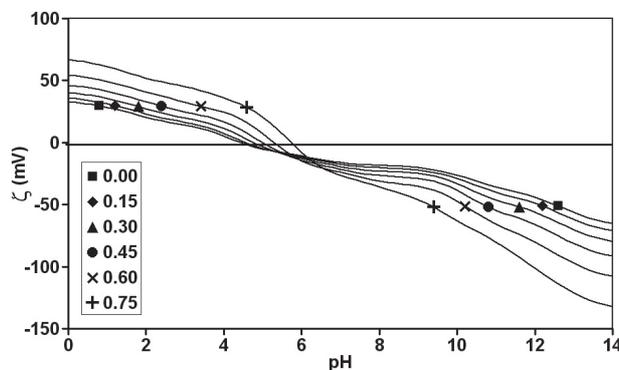
**Table IV.** Intercept (A), slope (B), standard error (Err) and correlation coefficient ( $R^2$ ) from Equation (5) expressing the relations between the absolute values of the electrostatic surface potential ( $\psi_0$  in mV) and the reciprocal dielectric constant ( $1/\epsilon$ ) for the lines  $\text{pH}_0$ ,  $\text{pH}_{1,50\%}$  and  $\text{pH}_{2,50\%}$  from Figure 3, and the isolines  $\text{pH}_{1,10\%}$ ,  $\text{pH}_{2,10\%}$ ,  $\text{pH}_{1,90\%}$  and  $\text{pH}_{2,90\%}$  from Figure 11, corresponding to a casein solubility equal to 10% and 90% for the pH range below (1) and over (2) the minimal solubility point ( $\text{pH}_0$ ). N = number of experimental points.

	A	B	Err	$R^2$	N
$\psi_0$	$-6.4 \pm 2.9$	$1475 \pm 164$	1.7	0.953	6
$\psi_{1,2,50\%}$	$-54 \pm 7.6$	$7034 \pm 429$	6.3	0.964	12
$\psi_{1,2,10\%}$	$-71 \pm 6.5$	$6645 \pm 364$	5.3	0.710	12
$\psi_{1,2,90\%}$	$-43 \pm 5.3$	$7192 \pm 300$	4.4	0.983	12
$\psi_{1,2,90\%-10\%}$	$29 \pm 4.9$	$547 \pm 278$	4.1	0.279	12

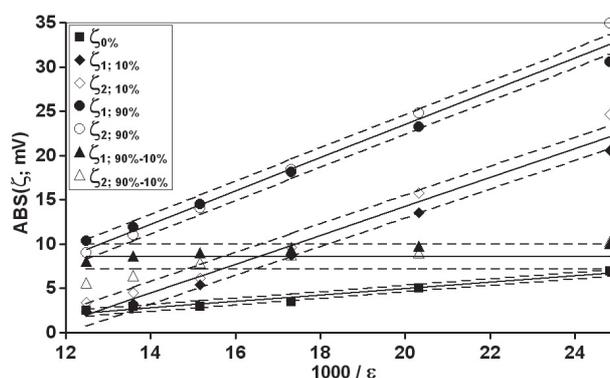
The Debye length is the distance from the surface of the protein molecule, over which the electrostatic surface potential drops to  $\exp(-1) = 0.368$  of its initial level  $\psi_0$ . For the experimental conditions applied in this work, the nominal thickness of the electric double layer varied between 0.2 and 0.3 nm at very low or very high pH levels and 3.0 and 4.3 nm near the isoelectric points (Fig. 12). This means that  $\lambda_D$  varied between 5% and 95% of the average radius of casein monomers. For the same pH level,  $\lambda_D$  decreased progressively with the rise in the ethanol volume fraction. For casein micelles in milk, the Debye length is  $\sim 1$  nm [40, 75].

The casein monomer zeta potential varied between +33 and +67 mV at pH 0 and -65 and -132 mV at pH 14 (Fig. 13). As

with the electrostatic surface potential, the absolute value of the zeta potential decreased with drops in the ethanol volume fraction. Within the 4.5 to 6.6 pH range, the effect of the ethanol volume fraction is opposite to that  $4.5 > \text{pH} > 6.6$ . For milk pH, the zeta potential decreased from -16 mV for water solution to -26 mV for 75% ethanol solution. The zeta potential of casein micelles in milk and in different buffers varied between -30 and -10 mV [2, 10, 12, 20, 23, 25, 26, 49, 52, 53]. In absolute values, the zeta potential is lower for smaller micelles [51] and also decreases with the addition of calcium, magnesium, copper and iron ions [53], and when the pH is lowered [2, 13]. When the ionic strength is high, the zeta potential may be much



**Figure 13.** Evolution of the zeta potential ( $\zeta$ ) of casein monomers, as a function of pH and ethanol volume fraction (0 to 0.75), calculated by Equation (10) for  $T = 293$  K.



**Figure 14.** Absolute value of the zeta potential ( $\zeta$ ) of casein monomers as a function of the reciprocal relative dielectric constant ( $1/\epsilon$ ) for the line  $\text{pH}_0$  ( $\zeta_{0\%}$ ), from Figure 3 and the isolines  $\text{pH}_{1:10\%}$ ,  $\text{pH}_{2:10\%}$ ,  $\text{pH}_{1:90\%}$  and  $\text{pH}_{2:90\%}$  from Figure 4, corresponding to casein solubilities of 10% and 90% for the pH range below ( $_1$ ) and over ( $_2$ ) the minimal zeta potential ( $\zeta$ ). Solid lines represent Equation (5) with coefficients A and B given in Table V and broken lines show standard deviation limits.

smaller than the electrostatic surface potential [10, 52, 53].

The zeta potential, corresponding to the isoline  $\text{pH}_0$  from Figure 3, increased from +2.5 mV for water solutions to +7.0 mV for ethanol volume fractions equal to 0.75 (Fig. 14, Tab. V). The absolute values of the zeta potentials  $\zeta_{1:10\%}$ ,  $\zeta_{2:10\%}$ ,  $\zeta_{1:90\%}$ , and  $\zeta_{2:90\%}$ , corresponding to casein solubilities of 10% and 90% for the pH range below ( $_1$ ) and over ( $_2$ ) the minimal pH solubility, are inversely proportional to the relative dielectric constant. Within the limits of stand-

ard error ( $\pm 1.5$  mV), the zeta potential levels are similar for the corresponding isolines on both sides of the minimal casein solubility or minimal surface potential. The zeta potential difference between 90% and 10% casein solubility is on average  $8.6 \pm 1.4$  mV, independently of the dielectric constant of the solvent. This means that on both sides of the isoelectric point, an increase in the zeta potential of about 9 mV raises the casein solubility from 10% to 90%.

Protein molecules in solution are also subjected to van der Waals attractive interactions,

**Table V.** Intercept (A), slope (B), standard error (Err) and correlation coefficient ( $R^2$ ) from Equation (5) expressing the relations between the absolute values of the zeta potential ( $\zeta$  in mV) and the reciprocal dielectric constant ( $1/\epsilon$ ) for the lines  $\text{pH}_0$ ,  $\text{pH}_{1;50\%}$  and  $\text{pH}_{2;50\%}$  from Figure 3, and the isolines  $\text{pH}_{1;10\%}$ ,  $\text{pH}_{2;10\%}$ ,  $\text{pH}_{1;90\%}$  and  $\text{pH}_{2;90\%}$  from Figure 14, corresponding to a casein solubility equal to 10% and 90% for the pH range below (<sub>1</sub>) and over (<sub>2</sub>) the minimal solubility point ( $\text{pH}_0$ ). N = number of experimental points.

	A	B	Err	$R^2$	N
$\zeta_0$	$-2.3 \pm 0.7$	$362 \pm 37$	0.4	0.960	6
$\zeta_{1,2;50\%}$	$-16 \pm 1.9$	$1798 \pm 105$	1.5	0.967	12
$\zeta_{1,2;10\%}$	$-18 \pm 1.6$	$1627 \pm 88$	1.3	0.972	12
$\zeta_{1,2;90\%}$	$-14 \pm 1.4$	$1878 \pm 76$	1.1	0.984	12
$\zeta_{1,2;90\%-10\%}$	$4 \pm 1.1$	$251 \pm 62$	0.9	0.618	12

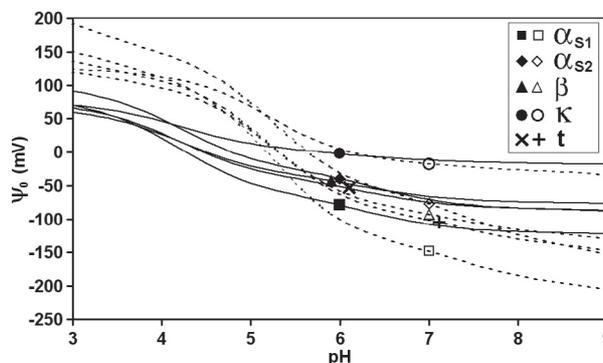
which can partly or totally counterbalance the electrostatic repulsion. The Hamaker constant ( $A_H$ ) has to be known to determine the van der Waals interaction energy. However, the Hamaker constant has been determined for only very few proteins [24, 60]. Probably none of the casein family has been studied so far. For the same protein, depending on the method and the theoretical approach used, the Hamaker constant can vary between 2 and 10  $k_B T$  [24]. For casein micelles Kruif and Tuinier [40] applied  $A_H = 1 k_B T$ , assuming that the protein density of the casein micelles is 6 times lower than in globular proteins. Without having more precise evaluation of the Hamaker constant for caseins, it seems premature to estimate correctly the van der Waals attractive energy.

In this work we have presented the electrical charge ( $z$ ), the electrostatic surface potential ( $\psi_0$ ) and the zeta potential of hypothetical molecules with a weight average amino acid composition of four basic casein fractions  $\alpha_{S1}$ -,  $\alpha_{S2}$ -,  $\beta$ - and  $\kappa$ -. The weight average molecular weight of this virtual, total casein would be 23.38  $\text{kg}\cdot\text{mol}^{-1}$ . This approach seems to work for comparing the solubility of total casein with the electrostatic surface potential or the zeta potential of such a hypothetical total casein molecule.

In reality, the four basic casein fractions in solution coexist and interact independently. Their electrostatic surface potential ( $\psi_0$ ) as a function of pH is presented in Figure 15. At pH 3 in water solutions, it is  $\alpha_{S2}$ -casein that is the most positively charged

(+92 mV). At pH 6 to 9 it is  $\alpha_{S1}$ -casein that is most negatively charged (−122 mV at pH 9). So-called total casein ( $t$  in Fig. 15) evolves somewhere in the middle. In 60% (v/v) ethanol solutions, the amplitude of the electrostatic surface potential increases to +192 mV at pH 3 for  $\alpha_{S2}$ -casein and to −204 mV for  $\alpha_{S1}$ -casein at pH 9, but the order of fractions does not change significantly. The isoelectric point increases on average by 1 pH unit. The enhancement of the differences of the electrostatic potential between the casein fractions by ethanol could be employed for the preparation of enriched or purified  $\alpha_{S1}$ -casein [30, 78].

The evolution of the electrostatic surface potential ( $\psi_0$ ) of individual casein fractions, or the hypothetical total casein, explains casein solubility quite well as a function of pH and the dielectric constant of the solvent. It has to be stressed, however, that at  $\text{pH} < 1$  the molecules have only positive charges and at  $\text{pH} > 13.5$  they are almost exclusively negatively charged. Within the pH range between 3 and 9, analysed in this work, the casein molecules have both sorts of charges. For this reason, not only repulsive but also attractive electrostatic interactions exist within the same molecules and between different casein molecules, leading to the formation of casein micelles [19, 22, 61, 62]. The state of casein aggregation in sodium caseinate solutions is not very well known. Dalglish and Law [11] found particle presence of around 100 nm in water solutions containing 15  $\text{mmol}\cdot\text{L}^{-1}$  of  $\text{Ca}^{2+}$  for spray-dried, 50 nm



**Figure 15.** Evolution of the electrostatic surface potential ( $\psi_0$ ) of principal casein fractions ( $\alpha_{S1}$ -,  $\alpha_{S2}$ -,  $\beta$ - and  $\kappa$ -) dissolved in water (solid lines) or in 60% (v/v) ethanol (broken lines) as a function of pH. The lines (t) represent total casein.

for roller-dried and 300 nm for laboratory prepared and freeze-dried sodium caseinate samples. For industrial calcium caseinates dissolved in water, Moughal et al. [44] found the particle size distribution to be bimodal, with the radius of the first peak of around 170–200 nm and that of the second peak of around 13–17  $\mu\text{m}$ . In the absence of calcium ions, the aggregates in the sodium caseinate are too small to be detected by laser methods based on the intensity of the scattered light [11, 44, 74].

#### 4. CONCLUSION

Casein solubility is a function of the intermolecular repulsion forces governed by the ionisation process of acidic and amino groups, being a function of pH and of the solvent's dielectric constant.

Equations describing the ionisation phenomenon, the electrostatic surface potential and the electrostatic surface energy quite satisfactorily represent casein solubility in water and in water ethanol mixtures.

The same equations would be applicable for other proteins and other polar solvents such as, for example, alcohols, amides, amines, esters, glycols, ketones, nitriles and sulfoxides.

A similar approach could be applied to studying the solubility of individual casein

fractions and especially of  $\alpha_{S1}$ -casein in neutral and slightly basic solutions and  $\alpha_{S2}$ -casein in acidic conditions.

#### REFERENCES

- [1] Åkerlöf G., Dielectric constants of some organic solvent-water mixtures at various temperatures, *J. Amer. Chem. Soc.* 54 (1932) 4125–4139.
- [2] Anema S.G., Klostermeyer H.,  $\zeta$ -Potentials of casein micelles from reconstituted skim milk heated at 120 °C, *Int. Dairy J.* 6 (1966) 673–687.
- [3] Banks W., Muir D.D., Effect of alcohol content on emulsion stability of cream liqueurs, *Food Chem.* 18 (1985) 139–152.
- [4] Banks W., Muir D.D., Wilson A.G., Extension of the shelf life of cream-based liqueurs at high ambient temperatures, *J. Food Technol.* 16 (1981) 587–595.
- [5] Banks W., Muir D.D., Wilson A.G., The formulation of cream-based liqueurs, *Milk Ind.* 83 (1981) 16,18.
- [6] Banks W., Muir D.D., Wilson A.G., Formulation of cream-based liqueurs: a comparison of sucrose and sorbitol as the carbohydrate component, *J. Soc. Dairy Technol.* 35 (2) (1982) 41–43.
- [7] Bingham E.W., Influence of temperature and pH on the solubility of  $\alpha_{S1}$ ,  $\beta$  and  $\kappa$ -casein, *J. Dairy Sci.* 54 (1971) 1077–1080.
- [8] Chavez M.S., Negri L.M., Taverna M.A., Cuartín A., Bovine milk composition parameters

- affecting the ethanol stability, *J. Dairy Res.* 71 (2004) 201–206.
- [9] Clark D.C., Smith L.J., Influence of alcohol-containing spreading solvents on the secondary structure of proteins: A circular dichroism investigation, *J. Agric. Food Chem.* 37 (1989) 627–633.
- [10] Dalgleish D.G., Measurement of electrophoretic mobilities and zeta-potentials of particles from milk using laser Doppler electrophoresis, *J. Dairy Res.* 51 (1984) 425–438.
- [11] Dalgleish D.G., Law A.J.R., Sodium caseinates - composition and properties of different preparations, *J. Soc. Dairy Technol.* 41 (1988) 1–4.
- [12] Darling D.F., Dickson J., The determination of the zeta potential of casein micelles, *J. Dairy Res.* 46 (1979) 329–332.
- [13] Darling D.F., Dickson J., Electrophoretic mobility of casein micelles, *J. Dairy Res.* 46 (1979) 441–461.
- [14] Davies D.T., White J.C.D., The relation between the chemical composition of milk and the stability of the caseinate complex. II. Coagulation by ethanol, *J. Dairy Res.* 25 (1958) 256–266.
- [15] Donnelly W.J., Ethanol stability of casein solutions as related to storage stability of dairy-based alcoholic beverages, *J. Food Sci.* 52 (1987) 389–393.
- [16] Downey W.K., Murphy R.F., The temperature-dependent dissociation of  $\beta$ -casein from bovine casein micelles and complexes, *J. Dairy Res.* 37 (1970) 361–372.
- [17] Dufour E., Bertrand-Harb C., Haertlé T., Reversible effects of medium dielectric constant on structural transformation of  $\beta$ -lactoglobulin and its retinal binding, *Biopolymers* 33 (1993) 589–598.
- [18] Dufour E., Robert P., Bertrand-Harb C., Haertlé T., Conformation change of  $\beta$ -lactoglobulin: An ATR infrared spectroscopy study of the effect of pH and ethanol, *J. Protein Chem.* 13 (1994) 143–149.
- [19] Evans M.T.A., Irons L., Jones M., Physicochemical properties of  $\beta$ -casein and some carboxyacyl derivatives, *Biochim. Biophys. Acta* 229 (1971) 411–422.
- [20] Famelart M.H., Hardy M.H., Brulé G., Étude des facteurs d'extraction de la caséine- $\beta$ , *Lait* 69 (1989) 47–57.
- [21] Famelart M.H., Lepesant F., Gaucheron F., Le Graët Y., Schuck P., pH-induced physicochemical modifications of native phosphocaseinate suspensions: Influence of aqueous phase, *Lait* 76 (1996) 445–460.
- [22] Farrer D., Lips A., On the self-assembly of sodium caseinate, *Int. Dairy J.* 9 (1999) 281–286.
- [23] Green M.L., Crutchfield G., Density-gradient electrophoresis of native and of rennet-treated casein micelles, *J. Dairy Res.* 38 (1971) 151–164.
- [24] Gripon C., Legrand L., Rosenman I., Vidal O., Robert M.C., Boué F., Lysozyme-lysozyme interactions in under- and super-saturated solutions: a simple relation between the second virial coefficients in H<sub>2</sub>O and D<sub>2</sub>O, *J. Crystal Growth* 178 (1997) 575–584.
- [25] Guillaume C., Jiménez L., Cuq J.L., Marchesseau S., An original pH-reversible treatment of milk to improve rennet gelation, *Int. Dairy J.* 14 (2004) 305–311.
- [26] Guillaume C., Gastaldi E., Cuq J.L., Marchesseau S., Effect of pH on rennet clotting properties of CO<sub>2</sub>-acidified skim milk, *Int. Dairy J.* 14 (2004) 437–443.
- [27] Harvey A.H., Prausnitz J.M., Dielectric constants of fluid mixtures over a wide range of temperature and density, *J. Solution Chem.* 16 (1987) 857–869.
- [28] Hewedi M.M., Mulvihill D.M., Fox P.F., Recovery of milk protein by ethanol precipitation, *Ir. J. Food Sci. Technol.* 9 (1985) 11–23.
- [29] Hiemenz P.C., Rajagopalan R., Principles of Colloid and Surface Chemistry, 3rd edn., Marcel Dekker Inc., New York, USA, 1997.
- [30] Hipp N.J., Groves M.L., Custer J.H., McMeekin T.L., Separation of  $\alpha$ ,  $\beta$  and  $\gamma$  casein, *J. Dairy Sci.* 35 (1952) 272–281.
- [31] Horne D.S., Ethanol stability of casein micelles - a hypothesis concerning the role of calcium phosphate, *J. Dairy Res.* 54 (1987) 389–395.
- [32] Horne D.S., Ethanol stability, in : Fox P.F. (Ed.), *Advanced Dairy Chemistry*, vol.1 : Proteins, Elsevier Applied Science, London, UK, 1992, pp. 657–689.
- [33] Horne D.S., Parker T.G., The pH sensitivity of ethanol stability of individual cow milks, *Neth. Milk Dairy J.* 34 (1980) 126–130.
- [34] Horne D.S., Parker T.G., Factors affecting the ethanol stability of bovine milk. I. Effect of serum phase components, *J. Dairy Res.* 48 (1981) 273–284.
- [35] Horne D.S., Parker T.G., Factors affecting the ethanol stability of bovine milk. II. The origin of pH transition, *J. Dairy Res.* 48 (1981) 285–291.
- [36] Horne D.S., Parker T.G., Factors affecting the ethanol stability of bovine milk. V. Effect

- of chemical modification of milk proteins, *J. Dairy Res.* 49 (1982) 449–457.
- [37] IDF, Milk. Determination of non-caseinic nitrogen content. Standard 29, Int. Dairy Fed., Brussels, Belgium, 1964.
- [38] IDF, Milk. Determination of nitrogen content. Standard 20B, Int. Dairy F., Brussels, Belgium, 1993.
- [39] Igarashi Y., Separation of caseins by chemical procedures, *Int. Dairy J.* 9 (1999) 377–378.
- [40] Kruif C.G., Tuinier R., Colloidal Interactions. Stabilisation of food colloids by polymers, in: Dickinson E. (Ed.), *Food Colloids. Interactions, Microstructure and Processing*, The Royal Society of Chemistry, Cambridge, UK, 2005, pp. 61–73.
- [41] Lynch A.G., Mulvihill D.M., Effect of sodium caseinate on the stability of cream liqueurs, *Int. J. Dairy Technol.* 50 (1997) 1–7.
- [42] Maubois J.L., Léonil J., Peptides du lait à activité biologique, *Lait* 69 (1989) 245–269.
- [43] Mohammed K.S., Fox P.F., Heat and alcohol-induced coagulation of casein micelles, *Ir. J. Food Sci. Technol.* 10 (1986) 47–55.
- [44] Morr C.V., Functional properties of milk proteins and their use as food ingredients, in: Fox P.F. (Ed.), *Developments in dairy chemistry*, vol. 1: Proteins, Applied Science Publishers, London, UK, 1982, pp. 375–399.
- [45] Moughal K.I., Munro P.A., Singh H., Suspension stability and size distribution of particles in reconstituted, commercial calcium caseinates, *Int. Dairy J.* 10 (2000) 683–690.
- [46] Muir D.D., Cream liqueur manufacture—assessment of efficiency of methods using a viscometric technique, *Dairy Ind. Int.* 52 (1987) 38–40.
- [47] Muir D.D., Formulation of a cream liqueur model system for use in coffee, *N. Z. J. Dairy Sci. Technol.* 23 (1988) 1–9.
- [48] Muir D.D., Dalgleish D.G., Differences in behaviour of sodium caseinates in alcoholic media, *Milchwissenschaft* 42 (1987) 770–772.
- [49] O’Connell J.E., Fox P.F., Proposed mechanism for the effect of polyphenols on the heat stability of milk, *Int. Dairy J.* 9 (1999) 523–536.
- [50] O’Kennedy B.T., Cribbin M., Kelly P.M., Stability of sodium caseinate to ethanol, *Milchwissenschaft* 56 (2001) 681–684.
- [51] Park S.Y., Niki R., Sano Y., Size effects of casein micelles on rennet gels in the presence of  $\beta$ -lactoglobuline, *Int. Dairy J.* 9 (1999) 379–380.
- [52] Pearce K.N., Moving boundary electrophoresis of native and rennet-treated casein micelles, *J. Dairy Res.* 43 (1976) 27–36.
- [53] Philippe M., Le Graët Y., Gaucheron F., The effects of different cations on the physico-chemical characteristics of casein micelles, *Food Chem.* 90 (2005) 673–683.
- [54] Pierre A., Étude de la stabilité du lait à l’alcool. Solubilité du phosphate et du calcium du lait en présence d’éthanol, *Lait* 65 (1985) 201–212.
- [55] Pierre A., Milk stability in ethalonic solutions, *J. Dairy Res.* 56 (1989) 521–527.
- [56] Pierre A., Brulé G., Mineral and protein equilibria between the colloidal and soluble phases of milk at low temperature, *J. Dairy Res.* 48 (1981) 417–428.
- [57] Robitaille G., Britten M., Petitclerc D., Effect of a differential allelic expression of  $\kappa$ -casein gene on ethanol stability of bovine milk, *J. Dairy Res.* 68 (2001) 145–149.
- [58] Roefs S.P.F.M., Walstra P., Dalgleish D.G., Horne D.S., Preliminary note on the change in casein micelles caused by acidification, *Neth. Milk Dairy J.* 39 (1985) 119–122.
- [59] Rose D., Relation between micellar and serum casein in bovine milk, *J. Dairy Sci.* 51 (1968) 1897–1902.
- [60] Schaik H.M., Smit J.A.M., Determination of the osmotic second virial coefficient and the dimerization of  $\beta$ -lactoglobulin in aqueous solutions with added salt at the isoelectric point. *Physical Chemistry Chemical Physics* 2 (2000) 1537–1541.
- [61] Schmidt D.G., Association of caseins and casein micelle structure, in: Fox P.F. (Ed.), *Developments in dairy chemistry*, vol. 1, Applied Science Publishers, London, UK, 1982, pp. 61–82.
- [62] Schmidt D.G., Payens T.A.J., van Markwijk B.W., Brinkhuis J.A., On the subunit of  $\alpha_{S1}$ -casein, *Biochem. Biophys. Res. Comm.* 27 (1967) 448–455.
- [63] Silva S.V., Malcata F.X., Caseins as source of bioactive peptides. Review, *Int. Dairy J.* 15 (2005) 1–15.
- [64] Smith R.L., Jr., Lee S.B., Komori H., Arai K., Relative permittivity and dielectric relaxation in aqueous alcohol solutions, *Liq. Phase Equil.* 144 (1998) 315–322.
- [65] Sommer H.H., Binney T.H., A study of the factors that influence the coagulation of milk in the alcohol test, *J. Dairy Sci.* 6 (1923) 176–197.
- [66] Swaisgood H.E., Chemistry of the caseins, in: Fox P.F. (Ed.), *Advanced dairy chemistry*,

- vol.1, Proteins, Elsevier Science Publishers Ltd., London, UK, 1992, pp. 63–110.
- [67] Swaisgood H.E., Protein and amino acid composition of bovine milk, in: Jensen R.G. (Ed.), *Handbook of Milk Composition*, Academic Press, San Diego, USA, 1995, pp. 464–468.
- [68] Szereniewicz M., Kiwk A., Kietczewska K., Ethanol-induced changes in proteins and some mineral compounds of milk, *Polish J. Food Nutr. Sci.* 8/49 (1999) 27–38.
- [69] Towler C., Conversion of casein curd to sodium caseinate, *N. Z. J. Dairy Sci. Technol.* 11 (1976) 24–29.
- [70] Towler C., Roller-dried sodium caseinate, *N. Z. J. Dairy Sci. Technol.* 11 (1976) 140–141.
- [71] Towler C., The manufacture and reconstitution characteristics of granular sodium caseinate, *N. Z. J. Dairy Sci. Technol.* 13 (1978) 71–76.
- [72] Trémillon B., *Électrochimie analytique et réactions en solution*. Tome 1, Masson, Paris, France, 1993.
- [73] Vuilleumard J.C., Gauthier S., Paquin P., Les ingrédients à base de protéines laitières: Obtention, propriétés et utilisation, *Lait* 69 (1989) 323–351.
- [74] Walstra P., *Physical Chemistry of Foods*, Marcel Dekker Inc., New York, USA, 2003.
- [75] Walstra P., Jenness R., *Dairy Chemistry and Physics*, John Wiley and Sons, New York, USA, 1984.
- [76] Zadow J.G., The rate of addition of alcohol has a major effect on the alcohol stability of skim milk, *Aust. J. Dairy Technol.* 48 (1993) 38–39.
- [77] Zadow J.G., Alcohol-mediated temperature-induced reversible dissociation of the casein micelle in milk, *Aust. J. Dairy Technol.* 48 (1993) 78–81.
- [78] Zittle C.A., Custer J.H., Purification and some of the properties of  $\alpha_s$ -casein and  $\kappa$ -casein, *J. Dairy Sci.* 46 (1963) 1183–1188.
- [79] Zittle C.A., Pepper L., Influence of hydrogen and calcium ion concentrations, temperature, and other factors on the rate of aggregation of casein, *J. Dairy Sci.* 41 (1958) 1671–1682.
- [80] Zittle C.A., Cerbulis J., Pepper L., Della Monica E.S., Preparation of calcium-sensitive  $\alpha$ -casein, *J. Dairy Sci.* 42 (1959) 1897–1902.